

## (-)-EPIAFZELECHIN 5-O-β-D-GLUCOSIDE FROM *CRATAEVA RELIGIOSA*

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**Key Word Index**—*Crataeva religiosa*, Cappariaceae, flavan-3-ols, (-)-epiafzelechin 5-O-β-D-glucoside

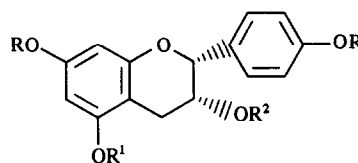
**Abstract**—Epiafzelechin 5-glucoside was characterized from the bark of *Crataeva religiosa* together with other known compounds

*Crataeva religiosa* Forst f is well known for its medicinal uses [1] Earlier chemical work on this plant is confined to triterpenoids and flavanoids [2-4] In the present paper we report the isolation and characterization of flavan-3-ols and a glucoside from the alcoholic extract of this plant Flavan-3-ol glucosides are of very rare occurrence in the plant kingdom and mostly occur as gallates [5] Previous reports of such glycosides are restricted to a few flavan-3-ols [6] The high percentage of (-)-epiafzelechin 5-glucoside in *C. religiosa* is most unusual

The defatted alcoholic extract of the plant after concentration and solvent extraction with ethyl acetate resulted in the separation of four compounds after column chromatography over silica gel Three of these were characterized as sitosterol glucoside, (-)-epiafzelechin and (-)-catechin The last and the most polar of the four compounds which is being reported for the first time was crystallized from ethyl acetate and was identified as (-)-epiafzelechin 5-O-β-D-glucoside (**1**) in the following manner Elemental analysis corresponded to the molecular formula C<sub>21</sub>H<sub>24</sub>O<sub>10</sub> IR showed very strong absorption at 3500 cm<sup>-1</sup> and no absorption for a carbonyl function Its UV spectrum in methanol showed maxima at 278 and 315 nm characteristic of a flavone system No UV shift with AlCl<sub>3</sub> indicated the absence of an ortho-dihydroxy grouping In the <sup>1</sup>H NMR spectrum of the compound in DMSO-*d*<sub>6</sub> two doublets at δ 7.40 and 6.83 (*J* = 8.5 Hz) were assigned to the protons of an A<sub>2</sub>B<sub>2</sub> system in ring B Two *meta* coupled doublets at δ 6.10 and 6.23 (*J* = 2.5 Hz) are due to C-6 and C-8 protons and C-4 protons signals were observed in the form of an envelope at δ 2.90 Acetylation of **1** gave a hepta-acetate **1a** Acetate signals were visible at δ 1.90, 2.0 and 2.30 for C-3 acetate, sugar acetoxy and aromatic acetate groups, respectively The signals for A<sub>2</sub>B<sub>2</sub> protons shifted to δ 7.03 and 7.40, respectively The C-6 and C-8 protons were located at δ 6.36 and 6.46 These two doublets however, merged to become a two proton singlet when the NMR spectrum was run in CCl<sub>4</sub> with a few drops of CDCl<sub>3</sub> added to fully solubilize the substance

Hydrolysis of **1** with emulsin gave glucose and an aglycone **1b** In the <sup>1</sup>H NMR spectrum of **1b** in DMSO-*d*<sub>6</sub>, A<sub>2</sub>B<sub>2</sub> proton signals were observed at δ 6.93 and 7.30 and signals for two *meta* coupled protons of ring A were located at δ 5.83 and 5.96 Also, a weakly coupled singlet at δ 4.96 was assigned to the C-2 proton Acetylation of **1b** gave a tetra-acetate **1c** In the <sup>1</sup>H NMR spectrum of **1c**

three extra signals were visible at δ 2.80 for two protons at C-4, 5.03 and 5.30 each integrating for a single proton assignable to the C-2 and C-3 protons, respectively The weak coupling between these two is indicative of the *cis* nature of these protons Compounds **1b** and **1c** were confirmed as epiafzelechin and its acetate by melting point, specific rotation and spectral data The shift of C-6 and C-8 protons to δ 6.43 and 6.63 and A<sub>2</sub>B<sub>2</sub> protons of the B ring to δ 7.0 and 7.30 clearly shows glucosidation in the A ring



- 1** R = R<sup>2</sup> = H  
 R<sup>1</sup> = Glc  
**1a** R = R<sup>2</sup> = COMe  
 R<sup>1</sup> = Glc Tetra Acetate  
**1b** R = R<sup>1</sup> = R<sup>2</sup> = H  
**1c** R = R<sup>1</sup> = R<sup>2</sup> = COMe  
**1d** R = Me, R<sup>1</sup> = R<sup>2</sup> = H  
**1e** R = Me, R<sup>1</sup> = R<sup>2</sup> = COMe

Table 1 <sup>13</sup>C NMR of (-)-epiafzelechin 5-glucoside

C-2	81.2	C-2'	127.8
C-3	64.7	C-3'	114.5
C-4	27.9	C-4'	154.9
C-4a	99.5	C-5'	114.5
C-5	156.5	C-6'	127.8
C-6	96.0	C-1''	100.7
C-7	156.1	C-2''	73.8
C-8	96.9	C-3''	76.8
C-8a	156.1	C-4''	70.1
		C-5''	78.3
C-1'	129.5	C-6''	61.2

Methylation of **1** and subsequent hydrolysis of the methylated product gave **1d**. The  $^1\text{H}$  NMR spectrum of **1d** in  $\text{CDCl}_3$  displayed signals for C-3 and C-2 protons at  $\delta$  4.26 (*br s*) and  $\delta$  4.96 (*s*), respectively.  $\text{A}_2\text{B}_2$  proton signals were located at  $\delta$  6.90 and 7.40. The C-6 and C-8 proton signals were observed at  $\delta$  6.03 and 6.13, respectively, and signals for C-4 protons were distinguishable at  $\delta$  2.96. Acetylation of **1d** gave **1e**. In the  $^1\text{H}$  NMR spectrum ( $\text{CDCl}_3$ ) of **1e** signals for C-6 and C-8 protons registered a shift to  $\delta$  6.30 and 6.36, respectively. The other signals remained almost at their original positions. This observation is also indicative of the glucosidation in the A ring.

The position of glucose in the A ring was finally established at C-5 from the Gibb's test of **1** and **1d** which was positive in the latter case. The position was also corroborated from the comparative study of C-6 and C-8 protons shifts in the  $^1\text{H}$  NMR spectra of **1**, **1a** and **1e**.

#### EXPERIMENTAL

All the mps are uncorr. The  $^1\text{H}$  NMR spectra were recorded on a Varian T-60A and  $^{13}\text{C}$  NMR on a JEOL-90 with TMS as int standard. The plant material was procured from the Bombay region (R R L Herbarium accession number 1074) and the bark was extracted with petrol and then EtOH. The conc EtOH extract was subjected to partition with  $\text{C}_6\text{H}_6$ , EtOAc and *n*-BuOH, respectively. The EtOAc extract was concd, charged over a silica gel column and eluted with various proportions of  $\text{CHCl}_3$  and MeOH. Four pure compounds were obtained in this manner. The first three were identified as sitosterol glucoside, (-)-epiafzelechin and (-)-catechin by co-TLC, superimposable IR and comparison of  $^1\text{H}$  NMR spectra with authentic samples. The fourth and major component **1** was crystallized from EtOAc as buff coloured crystals, mp  $190^\circ$ ,  $[\alpha]_{\text{D}}^{\text{MeOH}} -38.3^\circ$ . It analysed for  $\text{C}_{21}\text{H}_{24}\text{O}_{10}$  (Found C, 57.76, H, 5.61%, calc C, 57.8, H, 5.5%).  $^1\text{H}$  NMR (DMSO-*d*<sub>6</sub>)  $\delta$  2.90 (envelope 2H, H-4), 6.10 (1H, *d*,  $J = 2.5$  Hz, H-6), 6.23 (1H, *d*,  $J = 2.5$  Hz, H-8), 6.83 (2H, *d*,  $J = 8.5$  Hz, H-3' and H-5'), 7.40 (2H, *d*,  $J = 8.5$  Hz, H-2' and H-4'). MS of **1** showed fragments at  $m/z$  436  $[\text{M}]^+$ , 396, 308, 274, 255, 254, 211, 139, 136, 98, 70, 61 (100%) and 44. IR  $\nu_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$  3400, 1610, 1530, 1500, 1450, 1370, 1220, 1110, 1050.

**Acetylation of 1** Acetylation ( $\text{Ac}_2\text{O}$ -pyridine) gave a hepta-acetate **1a** crystallized from MeOH, mp  $210^\circ$ .  $^1\text{H}$  NMR ( $\text{CDCl}_3$ )  $\delta$  1.90 (3H, *s*, OAc), 2.00 (12H,  $4 \times$  OAc), 2.30 (3H, *s*,  $2 \times$  ArOAc), 2.80 (2H, envelope, H-4), 3.76 (*br m*, -CH, sugar), 4.16 (2H, envelope  $\text{CH}_2$ , OAc sugar), 5.0-5.40 (6H, *m*, sugar and H-3 and H-4), 6.36 and 6.46 ( $2 \times$  1H, *d*,  $J = 2.5$  Hz, H-6, H-8), 7.03 (2H, *d*,  $J$

$= 8.5$  Hz, H-3', H-5'), 7.40 (2H, *d*,  $J = 8.5$  Hz, H-2', H-6')

**Enzyme hydrolysis of 1** Compound **1** (100 mg) was dissolved in EtOH and emulsin added. The mixture was kept at room temp for 24 hr and processed as usual. The sugar was identified as glucose by PC. The aglycone **1b** was extracted with EtOAc and crystallized from the same solvent mp  $246^\circ$ ,  $[\alpha]_{\text{D}}^{\text{MeOH}} -63^\circ$ . It analysed for  $\text{C}_{15}\text{H}_{14}\text{O}_5$  (Found C, 65.75, H, 5.04, Calc C, 65.69, H, 5.11%).  $^1\text{H}$  NMR of **1b** (DMSO-*d*<sub>6</sub>)  $\delta$  2.80 (2H, envelope, H-4), 4.96 (1H, *s*, H-2), 5.83 and 5.96 ( $2 \times$  1H, *d*,  $J = 2.5$  Hz, H-6, H-8), 6.93 (2H, *d*,  $J = 8.5$  Hz, H-3', H-5'), 7.30 (2H, *d*,  $J = 8.5$  Hz, H-2', H-6'). MS of **1b** showed fragments at  $m/z$  274  $[\text{M}]^+$ , 211, 167, 139 (100%), 136, 107, 51, 43. Acetylation of **1b** in pyridine and  $\text{Ac}_2\text{O}$  resulted in the formation of a tetra-acetate **1c** crystallized from MeOH, mp  $125-126^\circ$ .  $^1\text{H}$  NMR of **1c** ( $\text{CDCl}_3$ )  $\delta$  1.83 (3H, *s*, OAc), 2.26 (9H, *s*,  $3 \times$  Ar-OAc), 2.80 (2H, envelope, H-4), 5.03 (1H, *s*, H-2), 5.30 (1H, *m*, H-3), 6.43 (1H, *d*,  $J = 2.5$  Hz, H-6), 6.63 (1H, *d*,  $J = 2.5$  Hz, H-8), 7.00 (2H, *d*,  $J = 8.5$  Hz, H-3', H-5') and 7.30 (2H, *d*,  $J = 8.5$  Hz, H-2', H-6').

Methylation of **1** with  $\text{CH}_3\text{N}_2$  in MeOH gave a methyl ether which was hydrolysed with emulsin to give **1d**, an amorphous powder.  $^1\text{H}$  NMR of **1d** ( $\text{CDCl}_3$ )  $\delta$  2.96 (2H, envelope, H-4), 3.76 (3H, *s*, Ar-Ome), 3.86 (3H, *s*, Ar-Ome), 4.26 (1H, *br s*, H-3), 4.96 (1H, *s*, H-2), 6.03 and 6.13 ( $2 \times$  1H, *d*,  $J = 2.5$  Hz, H-6, H-8), 6.93 (2H, *d*,  $J = 8.5$  Hz, H-3', H-5'), 7.50 (2H, *d*,  $J = 8.5$  Hz, H-2', H-6'). Acetylation of **1d** (pyridine- $\text{Ac}_2\text{O}$ ) gave a diacetate **1e**, an amorphous powder.  $^1\text{H}$  NMR of **1e** ( $\text{CDCl}_3$ )  $\delta$  1.80 (3H, *s*, OAc), 2.20 (3H, *s*, Ar-OAc), 2.76 (2H, envelope, H-4), 3.70 (3H, *s*, Ar-Ome), 3.73 (3H, *s*, Ar-Ome), 5.00 (1H, *s*, H-2), 5.26 (1H, *m*, H-3), 6.30 and 6.36 ( $2 \times$  1H, *d*,  $J = 2.5$  Hz, H-6, H-8), 6.80 (2H, *d*,  $J = 8.5$  Hz, H-3', H-5'), 7.26 (2H, *d*,  $J = 8.5$  Hz, H-2', H-6').

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